

Octet RED96 Easy Operation Process

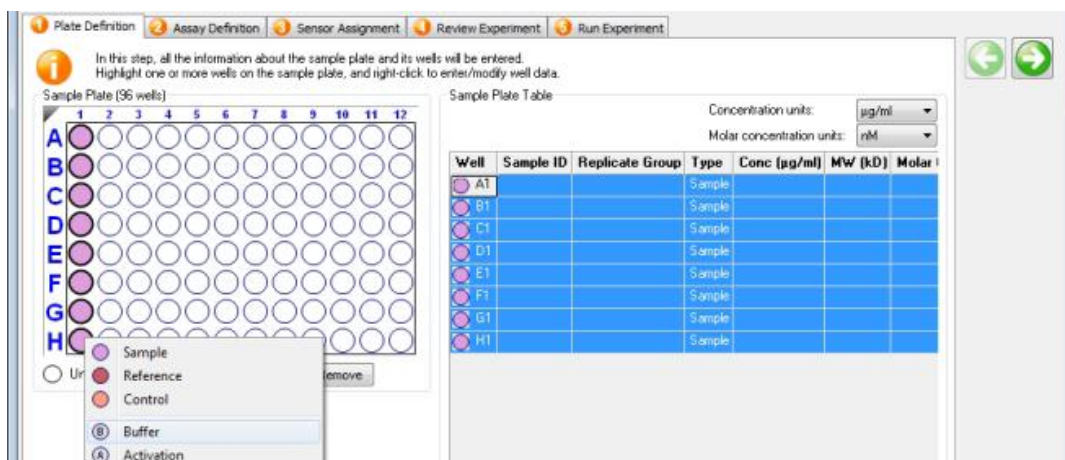
This process applies only to Octet RED96 assessment pass users. The instrument manager is not responsible for the behavior of using this simple procedure. It is strongly recommended to read Octet System Data Acquisition Software User Guide.

Start procedures :

1. Turn on the control computer, monitor and instrument power. (Instrument power needs to be turned on 40 minutes earlier)
2. Open Octet data acquisition software . (If the instrument fails to initialize, select Instrument: Reset to reestablish the connection)
3. Place the sensor tray and the 96 well plate at "right" place in the Octet Red96 and close the instrument door. (The instrument door opens only when the instrument status shows "ready")



4. Start basic kinetics or quantitation experiment . You can create a new experiment or modify the experiment you have used. (.fmf)
5. Set the program options in the order from left to right. ("Plate Definition", "Assay Definition", "Sensors Assignment", "Review Experiment", "Run Experiment")



6. The experimental data access directory is C: \ DATA.
7. When machine operation is running (yellow light), do not open the instrument door, so as not to affect the experimental data.

End procedures :

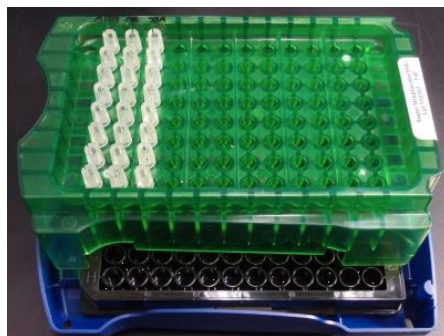
8. Save Log file to C: \ LOG.
9. Remove the sensor tray and 96 well plate, and turn off the control software and the computer screen
10. Instrument power remains on Monday to Friday, after the end of the experiment on Friday, turn off the computer and instrument power.

※Notice :

To burn the disc to access DATA, USB is not allowed to put into the host computer.

NOTES : (The following details, please check with Biosensors manufacturers)

- Biosensors should be soaked in hydration buffer for at least 30 minutes in sensor tray for conditioning and equilibration prior to usage. The hydration buffer is usually your experiment buffer except for some sensors (e.g. APS sensor) which might be water.



- Before start your big experiments, perform a non-specific binding test by using a blank sensor with your sample to check if there is binding of your sample to the sensor surface. If non-specific binding does occur, the buffer needs optimization. Adding 0.05% Tween20, 0.1mg/mL BSA or PEG400 may help. Otherwise, use different type of sensors.
- Please note that the following sensors may contain streptavidin: SA, SAX, AHC, AMC, Fab2G, NTA, His1K, HIS2, GST and CHO. If needed, ask for 10ug/mL biocytin to block unoccupied biotin binding sites on the sensor surface.
- It is better to design your experiments with double reference subtraction to eliminate background that is caused by buffer mismatch or insignificant non-specific binding.
- Buffers, reagents and samples (180 -220 μ L) are filled in a 96-well plate, which is placed on the temperature controlled orbital shaker sample stage.
- Please refer to [Octet Data Analysis Software 7.0 User Guide](#) for data analysis.